12. Management of the infected vascular access

Guideline 12.1. Infection of autogenous AV fistulae without fever or bacteraemia should be treated by appropriate antibiotics for at least 2 weeks (Evidence level III).

Guideline 12.2. Infection of autogenous AV fistulae with fever and/or bacteraemia should be treated by appropriate antibiotics given intravenously for 2 weeks. Excision of the fistula is required in case of infected thrombi and/or septic emboli (Evidence level IV).

Guideline 12.3. Infected graft AVFs should be treated by appropriate antibiotics given intravenously for 2 weeks and continued orally for 4 weeks. Depending on the presence of bacteraemia and/or infected thrombi segmental explantation of the graft with bypass needs to be considered (Evidence level III).

Guideline 12.4. Anastomotic infection is an indication for total graft explantation (Evidence level II).

Guideline 12.5. Catheter removal must be considered when catheter infection is suspected. Immediate removal should be performed in nontunnelled catheters when infection is diagnosed (Evidence level III).

Guideline 12.6. In tunnelled catheters with a short febrile and/or bacteraemic reaction, a delayed removal may be considered (Evidence level III). In septicaemia, immediate removal should be performed in tunnelled catheters as well.

AVF and prosthetic graft infection

Infection of autogenous AVF usually responds well to appropriate antibiotics given either orally or intravenously according to the presence of fever and/or bacteraemia. Surgical revision or excision of the fistula is required when infected thrombi, aneurysms and/or septic emboli are detected. Infection of graft AVFs is two to three times more frequent than autogenous AVFs [1]. Infection of the graft bears a worse prognosis and requires usually a surgical revision and/or explantation in addition to the antibiotic therapy. Salvaging prosthetic grafts may be attempted in certain circumstances. Several surgical techniques have been described in combination with antibiotic therapy. For localized abscesses, incision and drainage with graft preservation is needed. For more severe infection, such as infected thrombi, false

indicated. However, these salvaging techniques may be complicated because of local or generalized infection and sepsis. Therefore, in severe cases a complete explantation of all graft material with drainage is usually necessary.

Central venous catheter infection

Catheter-related infection is the major cause of morbidity in HD patients with central venous catheters [2-4]. Catheter infection is a potentially severe event that requires early diagnosis and appropriate management to prevent further complication. Diagnosis of catheter infection is relatively easy in symptomatic patients presenting with fever, pain, skin exit and/or track infection and bacteraemic episodes. It is much more difficult in silent catheter endoluminal contamination or low grade infection. In these cases, only specific blood and catheter clot culture will help to make the diagnosis [5]. Recently, it was shown that catheter clot culture after endoluminal brushing was more sensitive than blood culture to identify asymptomatic catheter infection (catheter contamination) [6,7]. Symptoms of infection includes chronic fever, bacteraemic episodes, catheter pain, inflammation of the exit site or tunnel. Infection of the catheter exit site or tunnel tract is usually observed by the dialysis nurse while clinical examination is performed at the time of dialysis connection. Silent contamination is suspected when recurrent febrile reactions during haemodialysis occur and bacterial pathogens (Staphylococcus aureus, S. epidermidis or other bacteria such as Gramnegatives) are identified in blood cultures. Catheterrelated septicaemia is usually associated with symptoms of endocarditis, arthritis, spondylarthritis or osteomyelitis.

Specific blood markers (leucocyte count and differentiation), C-reactive protein (CRP) and procalcitonin (PCT), help to diagnose early bacterial catheter infection. Catheter-related infection should be considered as a severe and potentially lethal complication. Prevention of infection should be a permanent preoccupation for care providers, that relies on hygienic measures [8] and strict protocols for handling catheters based on aseptic manipulation [9] and using specific dressings [10]. The regular and pre-emptive use of locking solutions (Citrate) with both antithrombotic and/or antiseptic properties has been confirmed to be effective in preventing catheter infection [11–14]. The topical application of antibiotic ointment on the skin exit site has proved to be efficient in reducing the incidence of bacteraemia at the expense of selecting antibiotic-resistant strains of bacteria [15–17]. The use of antibiotic-coated catheters or silver-treated catheters has been proposed to reduce the risk of infection, but conflicting results has been reported [18–20].

aneurysms, cellulitis, explantation of the infected graft segment and segmental bypass with a new graft is

Identification of patients at risk of infection is particularly important in diabetic patients and nasal

EBPG on vascular access

carriers of methicillin-resistant S. aureus (MRSA). In the latter patients, eradication of bacteria by means of topical antibiotic ointment has been associated with a significant reduction of bacteriaemias [21,22].

Catheter removal should be considered as the first line of treatment. Catheter withdrawal must be immediate when infection occurs in non-tunnelled catheters. Removal may be postponed for several days in tunnelled catheters. When this last option is applied the risk of septic complications of delayed catheter removal should be balanced with the benefits of keeping it in situ. This conservative option implies that the patient is regularly and carefully observed. In addition, the catheters should be disinfected by means of antimicrobial lock solutions and dissemination of the infection must be prevented by adequate systemic antibiotic therapy. When the catheter is left in place and in the absence of precise microbial information, antimicrobial therapy should include systemic antibiotic therapy effective against Staphylococcus species plus an adjunctive antimicrobial catheter lock. Antibiotic therapy is given for 2 weeks in order to sterilize all potential bacterial foci. Topical antibiotic therapy (catheter exit site) is initiated when there is associated local infection. Imaging techniques may help to diagnose catheterrelated infection. Ultrasound doppler methods can detect tunnel infection and/or subcutaneous abscesses along the catheter track. Phlebography and catheterography are indicated to diagnose infected thrombi located in the vein or fibrin sleeves surrounding the catheter tip. Isotopic imaging techniques using positron emission tomography (PET) may help to identify infected venous catheters and port devices [23].

Recommendations for further research

Improvement of needle design and education on strict aseptic cannulation techniques may possibly lower the incidence of infection in fistulae and grafts. Antibiotic-bonded grafts may possibly lower the incidence of graft infection. Newer catheter designs and locking solutions are important issues for further investigation of the prevention of central venous catheter-related infections.

References

- Kessler M, Hoen B, Mayeux D, Hestin D, Fontenaille C. Bacteremia in patients on chronic hemodialysis. A multicenter prospective survey. Nephron 1993; 64: 95–100
- Cheesbrough JS, Finch RG, Burden RP. A prospective study of the mechanisms of infection associated with hemodialysis catheters. J Infect Dis 1986; 154: 579–589
- Hoen B, Paul-Dauphin A, Hestin D, Kessler M. EPIBACDIAL: a multicenter prospective study of risk factors for bacteremia in chronic hemodialysis patients. J Am Soc Nephrol 1998; 9: 869–876
- Elseviers MM, Van Waeleghem JP. European Dialysis and Transplant Nurses Association/European Renal Care Association. Identifying vascular access complications among

- Maki DG, Weise CE, Sarafin HW. A semiquantitative culture method for identifying intravenous-catheter-related infection. N Engl J Med 1977; 296: 1305–1309
- McLure HA, Juste RN, Thomas ML, Soni N, Roberts AP, Azadian BS. Endoluminal brushing for detection of central venous catheter colonization–a comparison of daily vs. single brushing on removal. J Hosp Infect 1997; 36: 313–316
- Dobbins BM, Kite P, Catton JA, Wilcox MH, McMahon MJ. In situ endoluminal brushing: a safe technique for the diagnosis of catheter-related bloodstream infection. J Hosp Infect 2004; 58: 233–237
- Kaplowitz LG, Comstock JA, Landwehr DM, Dalton HP, Mayhall CG. A prospective study of infections in hemodialysis patients: patient hygiene and other risk factors for infection. Infect Control Hosp Epidemiol 1988; 9: 534–541
- Mermel LA, Farr BM, Sherertz RJ et al. Infectious Diseases Society of America; American College of Critical Care Medicine; Society for Healthcare Epidemiology of America. Guidelines for the management of intravascular catheter-related infections. Clin Infect Dis 2001; 32: 1249–1272
- European Best Practice Guidelines Expert Group on Hemodialysis, European Renal Association. Section VI. Haemodialysis-associated infection. Nephrol Dial Transplant 2002; 17[Suppl 7]: 72–87
- Betjes MG, van Agteren M. Prevention of dialysis catheterrelated sepsis with a citrate-taurolidine-containing lock solution. Nephrol Dial Transplant 2004;19: 1546–1551
- McIntyre CW, Hulme LJ, Taal M, Fluck RJ. Locking of tunneled hemodialysis catheters with gentamicin and heparin. Kidney Int 2004; 66: 801–805
- Weijmer MC, van den Dorpel MA, Van de Ven PJ et al. Randomized, clinical trial comparison of trisodium citrate 30% and heparin as catheter-locking solution in hemodialysis patients. J Am Soc Nephrol 2005; 16: 2769–2777
- Dogra GK, Herson H, Hutchison B et al. Prevention of tunneled hemodialysis catheter-related infections using catheter-restricted filling with gentamicin and citrate: a randomized controlled study. J Am Soc Nephrol 2002; 13: 2133–2139
- Maki DG, Stolz SS, Wheeler S, Mermel LA. A prospective, randomized trial of gauze and two polyurethane dressings for site care of pulmonary artery catheters: implications for catheter management. Crit Care Med 1994; 22: 1729–1737
- Levin A, Mason AJ, Jindal KK, Fong IW, Goldstein MB.
 Prevention of hemodialysis subclavian vein catheter infections by topical povidone-iodine. Kidney Int 1991; 40: 934–938
- 17. Sesso R, Barbosa D, Leme IL et al. Staphylococcus aureus prophylaxis in hemodialysis patients using central venous catheter: effect of mupirocin ointment. JAmSocNephrol1998; 9: 1085–1092
- Dahlberg PJ, Agger WA, Singer JR et al. Subclavian hemodialysis catheter infections: a prospective, randomized trial of an attachable silver-impregnated cuff for prevention of catheter-related infections. Infect Control Hosp Epidemiol 1995; 16: 506–511
- Trerotola SO, Johnson MS, Shah H et al. Tunneled hemodialysis catheters: use of a silver-coated catheter for prevention of infection—a randomized study. Radiology 1998; 207: 491–496
- Chatzinikolaou I, Finkel K, Hanna H et al. Antibiotic-coated hemodialysis catheters for the prevention of vascular catheterrelated infections: a prospective, randomized study. Am J Med 2003; 115: 352–357
- Johnson DW, MacGinley R, Kay TD et al. A randomized controlled trial of topical exit site mupirocin application in patients with tunnelled, cuffed haemodialysis catheters. Nephrol Dial Transplant 2002; 17: 1802–1807
- Lok CE, Stanley KE, Hux JE, Richardson R, Tobe SW, Conly J. Hemodialysis infection prevention with polysporin ointment. J Am Soc Nephrol 2003; 14: 169–179
- 23. Miceli MH, Jones Jackson LB, Walker RC, Talamo G, Barlogie B, Anaissie EJ. Diagnosis of infection of implantable

ESRD patients in Europe. A prospective, multicenter study. Nephrol News Issues 2003; 17: 61–64, 66–68, 99. central venous catheters by [18F]fluorodeoxyglucose positron emission tomography. Nucl Med Commun 2004; 25: 813–818